

Total Aflatoxin, Aflatoxin B₁ and Ochratoxin A Residues in Wheat Flour Produced in Kurdistan Province, Iran

Hiro Memari (MSc)

Department of Food Sciences and Technology, Islamic Azad University, Varamin-Pishva Branch, Varamin, Iran

Keiwan Ebrahimi Mohammadi (PhD)

Department of Food Sciences and Technology, Faculty of Agriculture, Islamic Azad University, Mahabad Branch, Mahabad, Iran

Peiman Esmailzadeh (MSc)

Department of Food Sciences and Technology, Faculty of Agriculture, Islamic Azad University, Mahabad Branch, Mahabad, Iran

Corresponding author: Keiwan Ebrahimi Mohammadi

E-mail: keiwan1976@yahoo.com

Tel: +989363679725

Address: Department of Food Sciences and Technology, Faculty of Agriculture, Islamic Azad University, Mahabad Branch, Mahabad, Iran

Received : 09 Jul 2016

Revised: 10 Aug 2016

Accepted: 15 Aug 2016

ABSTRACT

Background and objectives: Contamination of food products with mycotoxins is a public health problem. The International Agency for Research on Cancer has identified mycotoxins as hepatotoxic and carcinogenic agents to humans (Group 1). The Kurdistan Province is the ninth largest producer of wheat in Iran. We aimed to determine the level of contamination with total aflatoxin (TAF), aflatoxin B₁ (AFB₁) and ochratoxin A (OTA) in 66 wheat samples randomly selected from 11 wheat flour factories in spring and summer.

Methods: The level of toxins was measured by microtiter plate enzyme-linked immunosorbent assay (ELISA) using a microtitre plate ELISA reader and total AF, AFB₁ and OTA commercial kits.

Results: Overall, the level of TAF and AFB in 16.67% of the samples exceeded the maximum tolerable limit set by the Institute of Standard and Industrial Research of Iran (ISIRI). However, the level of OTA contamination did not exceed the maximum tolerable limit set by the ISIRI. In addition, the level of TAF, AFB₁ and OTA exceeded the maximum tolerable limit set by the EU in 68.18, 90.91 and 36.36% of the samples, respectively. The level of contamination with these mycotoxins differed significantly in spring and summer ($P < 0.05$).

Conclusion: The level of mycotoxin contamination in wheat samples produced in the Kurdistan Province is alarmingly high and appropriate measures should be taken to eliminate the causes of this issue.

KEYWORDS: Aflatoxin, Aflatoxin B₁, Ochratoxin A, Wheat, ELISA.

INTRODUCTION

Humans are exposed to toxins through consumption of foodstuffs contaminated with fungi and fungal byproducts. Since it is difficult to prevent growth of fungi in food, such exposures could not be easily avoided. When Good Agricultural Practices are not adopted during cultivation and storage of wheat grains, filamentous fungi such as *Fusarium*, *Penicillium* and *Aspergillus* may develop and produce secondary metabolites called mycotoxins (1, 2). Aflatoxin (AF) is a naturally occurring carcinogenic substance, which is extremely toxic to humans (3). According to the International Agency for Research on Cancer, they are hepatotoxic and carcinogenic agents to humans (Group 1) and capable of inducing liver cancer and cirrhosis as well as reducing individuals' immune resistance, causing outbreaks of hepatitis B (4). Kurdistan Province is a mountainous region located in west of Iran with cold climate, although some areas may have a warmer climate. According to the official survey of Statistical Centre of Iran, Kurdistan Province is the ninth largest producer of wheat in Iran (5). Nevertheless, some problems such as traditional agriculture practices, improper storage and distribution systems result in wheat and wheat flour fungal contamination in the province. According to the Institute of Standards and Industrial Research of Iran (ISIRI), the maximum tolerable limit for total AF (TAF), AFB₁ and ochratoxin A (OTA) in wheat flour is 15, 5 and 5 µg/Kg, respectively (6). However, the EU Commission (EC) has set the maximum tolerable limit for TAF and AFB₁ in cereals and all products derived from cereals at 4 and 2 µg/Kg, respectively. In addition, the maximum tolerable limit for OTA in all products derived from cereals is 3 µg/Kg (6).

MATERIAL AND METHODS

This study was performed on 66 samples randomly collected from 11 wheat flour manufactures in the Kurdistan Province, Iran. Sampling was carried out in spring (33 samples) and summer (33 samples) to determine the effect of season on the level AF and OTA contamination. The sampling was carried out at mid-month from 14-Kg whole-wheat flour bags.

All samples were stored in plastic bags and kept in refrigerator at 4-6 °C. Microtitre plate ELISA reader (Bio-Tek Inst, ELX 800, USA) and total AF, AFB₁ and OTA measurement kits (Agrastrip, Romer Labs, Singapore Pte Ltd.) were used. The Agra Quant® ELISA AF (Art. No: COKAQ1000), Agra Quant® ELISA AFB₁ (Art. No: COKAQ8000), Agra Quant® ELISA Ochratoxin A (Art. No: COKAQ2000) kits were obtained from Romero Labs for quantitative analysis of AF residues in cereals and feedstuffs. The functional properties of the kits used in the study are shown in table 1. After extraction of mycotoxins, all well plates were placed in an ELISA microplate reader (Bio-Tek Inst, ELX800, USA) with a 450 nm filter. Estimated amount of mycotoxins in each sample was calculated by extrapolation from the standard curve.

RESULTS

The results showed that all samples were contaminated with TAF. In addition, 68.18% of the wheat flour samples contained 11.83 ± 3.29 µg/Kg TAF (4.01-19.65 µg/Kg), which exceeds the maximum tolerable limit set by the EC (Table 2).

Moreover, the amount of TAF present in 16.67% of the samples exceeded the maximum tolerable limit for TAF set by the ISIRI (15 µg/Kg) (8).

Table 1- The functional properties of the kits used in the study

Mycotoxin	Limit of detection (µg/Kg)	Limit of quantitation (µg/Kg)	Range of quantitation(µg/Kg)	Standards (µg/Kg)	Percent recovery	CV(%)
OT	1.9	2	2-40	0,2,5,20,40	-	-
AFB ₁	2	2	2-50	0,2,5,20,50	87-103	≤10%
TAF	3	3	4-40	0,4,10,20,40	-	-

All data were analyzed in SPSS (version 21) using independent t-test and ANOVA. To evaluate differences between means, Tukey's test was used.

Table 2- TAF levels in wheat flour samples tested in the study

TAF($\mu\text{g}/\text{kg}$)	Number of positive samples	Mean ($\mu\text{g}/\text{kg}$) \pm standard error	Percentage
$\leq 4^a$	21	3.3 \pm 0.3	31.82
4-15	34	9/11 \pm 1.5	51.51
$>15^b$	11	17/17 \pm 2.1	16.67

a: The maximum tolerable limit of TAF according to EC regulations
b: The maximum tolerable limit of TAF according to ISIRI regulations

Table 3- Total AFB₁ levels in wheat flour samples tested in the study

AFB ₁ ($\mu\text{g}/\text{kg}$)	Number of positive samples	Mean ($\mu\text{g}/\text{kg}$) \pm standard error	Percentage
$\leq 2^a$	6	6	9.09
2-5	49	3.3 \pm 0.87	74.24
$>5^b$	11	5.28 \pm 1.23	16.67

a: The maximum tolerable limit of AFB₁ according to EC regulations
b: The maximum tolerable limit of AFB₁ according to ISIRI regulations

As shown in table 3, the amount of AFB₁ in 9.09% of the samples was lower than the limit of detection and allowed limit set by the EC (2 $\mu\text{g}/\text{kg}$). In addition, the level of AFB₁ in 16.67% of the samples was higher than the maximum tolerable limit set by the ISIRI (5 $\mu\text{g}/\text{kg}$).

OTA is found predominantly in cereal grains, cereal products, legumes, oilseed, coffee beans and feedstuff (9).

In our study, the level of OTA was determined to $3.17 \pm 0.18 \mu\text{g}/\text{kg}$. While all samples were contaminated with OTA, the level of OTA in 63.63% of the samples was

lower than the tolerable limit set by the EC (3 $\mu\text{g}/\text{kg}$). However, level of OTA in none of the samples exceeded the tolerable limit set by the ISIRI (5 $\mu\text{g}/\text{kg}$).

Mycotoxin levels in spring and summer

There were statistically significant differences between the level of TAF, AFB₁ and OTA in samples collected in spring and summer (Table 4).

The level of TAF, AFB₁ and OTA in both seasons was within the tolerable limit set by the ISIRI. However, only the level of OTA in spring was lower than the tolerable limit set by the EC.

Table 4- Mean level of mycotoxins in the samples collected in spring and summer

Mycotoxin	Season	Number of samples	Mean level ($\mu\text{g}/\text{kg}$)	Standard deviation
TAF	Spring	33	11.01 ^{A*}	4.24
	Summer	33	10.83 ^B	2.90
AFB ₁	Spring	33	3.65 ^{A*}	2.28
	Summer	33	3.12 ^B	2.11
OTA	Spring	33	2.08 ^{A*}	1.08
	Summer	33	3.77 ^B	1.54

* Statistically significantly difference ($P < 0.05$)

DISCUSSION

Contamination of food products is a major problem in Iran (10-14). The most staple food in Iran is wheat, which is susceptible to fungal contamination during cultivation, harvest, transport and storage. In our study, the level of TAF was higher than that reported by previous studies in Iran. In our study, 2.54% of the

samples were contaminated with AF, while 2.54% and 3.39% of the samples were contaminated with AFB₁ and AFG₁, respectively. Zinedine et al. reported that the mean level of TAF and AFB₁ in wheat flour was 0.07 ng/g and 0.07 ng/g, respectively. They also reported that none of the samples

was contaminated with AFB₂, AFG₁ and AFG₂ (15). Abdullah et al. analyzed 83 wheat flour samples in Malaysia (16), and reported that 21.7% of the samples was contaminated with AF. In another study, Giray et al. analyzed 41 wheat samples in Turkey, and reported that 59% of the samples was contaminated with AFB₁ (42%), AFB₂ (12%), AFG₁ (37%) and AFG₂ (12%) (17). Aydin et al. assessed the level of TAF in 100 wheat flour samples, and concluded that 45% of them contained 0.005-14.0 µg/Kg AF. In addition, the level of TAF in 2% of the samples exceeded the maximum tolerable limit in Turkey (4 µg/kg) (18). A study in Iran showed that 3.1% and 7.4% of wheat flour sample were contaminated with AFB₁ in summer and winter, respectively, with levels exceeding the permissible limit (19). In another study, the level AFB₁ was reported to be 0.93 ng/g, while 6.5% of the samples contained AFB₁ levels higher than the maximum limit set by international regulations (20). A study in Croatia reported that the mean level of AFB₁ was 16.3 µg/Kg in 475 wheat grain samples (21). Ayalew et al. detected AFB₁ contamination in 8.8% of 352 cereal samples (wheat, barley, sorghum, and teff grass) (22). Although the highest level of AFB₁ found in our study was lower than that in studies of Giray et al., Halt and Ayalew et al., it was higher than the level reported by studies of Vagef and Mahmoudi in Iran (13, 17, 21, 22).

A study determined the concentration of OTA in wheat flour samples collected from Ahvaz using HPLC method. In the mentioned study, 93.75% of the samples were contaminated with OTA at mean concentration of 0.09 ng/g, which is lower than its maximum tolerable limit in wheat according to the ISIRI and WHO (23). Yazdanpanah et al. found no OTA contamination in 14 barley and nine corn samples (destined for animal feed) collected from the Golestan and Mazandaran provinces in north of Iran (24). However, they found one case of AF and OTA co-contamination. Mahmoudi et al. found that 24.3, 11.4 and 14.3 % of the wheat flour samples from the Mazandaran Province contained AFB₁, TAF and OTA levels exceeding the maximum tolerable limit, respectively (25).

In our study, detection of OTA in 33.3% of wheat flour samples is alarming. Study of Zinedine et al. in Morocco reported that 40% of wheat samples were contaminated with

OTA at mean concentration of 1.73 µg/Kg (15). In Ethiopia, Ayalew et al. reported that 23.4% of 107 wheat samples had OTA-contamination, and the highest level of OTA recorded was 66.0 µg/Kg (22). In Italy, Muscarella et al. reported that 95 durum wheat, 15 flour wheat, 80 maize and 85 barley samples were contaminated with OTA (26).

In our study, the level of AFB₁ contamination in 3.1% and 7.4% of samples collected in summer and winter was higher than the international permissible limit, respectively. AF level in winter was higher than in summer, and the highest frequency of AF contamination in winter and summer was related to AFB₂ (98%) and AFG₁ (51%), respectively. The difference between the mean levels of AF in the two seasons may be related to geographical and climatic characteristics as well as temperature and humidity. Agricultural practices, soil characteristics, type of wheat, sampling method, and wheat and flour storage conditions can affect the contamination level in flour (19). Baliukoniene et al. showed that storage conditions have a significant impact on levels of AF contamination (27). In summer, temperature and humidity are higher and more suitable for mycotoxin production. However, we showed that TAF levels in spring were higher than in summer. This could be due to duration of wheat storage and other adverse conditions. Based on the results of our study and other studies, it can be concluded that there is a significant relationship between humidity and the level of TAF (19). Basilio showed that fungal growth and toxin production are influenced by the interaction between fungi, host and environment (28). It is well known that the type of crop, weather pattern, temperature, humidity, water activity, level of oxygen, poor storage condition, insufficient drying and presence of insects or rodents can affect growth of *Aspergillus* species and AF production (29). Unfortunately, most of the wheat produced in the Kurdistan Province is stored in small or tradition silos, and transported at unsuitable conditions.

CONCLUSION

The results of this study show that all wheat flour samples produced in the Kurdistan Province are contaminated with different levels of mycotoxins. We found that the level of contamination is different in spring and

summer. Considering the difference between the standard limits for mycotoxins set by the ISRI and EC and the alarming rate of contamination in our samples, it is necessary to review and modify the existing standards in accordance with the international standards.

REFERENCES

- Gregori R, Meriggi P, Pietri A, Formenti S, Baccarini G, Battilani P. *Dynamics of fungi and related mycotoxins during cereal storage in silo bags*. Food Control. 2013; 30(1): 280-7.
- Mylona K, Sulyok M, Magan N. *Relationship between environmental factors, dry matter loss and mycotoxin levels in stored wheat and maize infected with Fusarium species*. Food Additives & Contaminants: Part A. 2012; 29(7): 1118-28. doi: 10.1080/19440049.2012.672340.
- Songsermsakul P, Razzazi-Fazeli E. *A review of recent trends in applications of liquid chromatography-mass spectrometry for determination of mycotoxins*. Journal of Liquid Chromatography & Related Technologies. 2008; 31(11-12): 1641-86. DOI: 10.1080/10826070802126395.
- World Health Organisation(WHO). *International Agency for Research on Cancer. Some traditional herbal medicines, some mycotoxins, naphthalene and styrene*. IARC Press, Lyon, France. 2002; 82: 193-210.
- Institute of Standards and Industrial Research of Iran (ISIRI). *Wheat flour; Specification and test methods*. 2010; No.103. Available from www.isiri.org.
- Commission E. *Commission Regulation (EC) No 466/2001 of 8 March 2001 setting maximum levels for certain contaminants in foodstuffs*. Brussels: European Commission. 2001.
- Institute of Standards and Industrial Research of Iran (ISIRI). *Sampling methods to official control of mycotoxin levels, Food and agriculture products*. 2009. Available from www.isiri.org.
- Institute of Standards and Industrial Research of Iran (ISIRI). *Food and Feed Mycotoxins-Maximum tolerated level*.NO:5925. 2002. Available from www.isiri.org.
- MacDonald S, Wilson P, Barnes K, Damant A, Massey R, Mortby E, Shepherd MJ. *Ochratoxin A in dried vine fruit: method development and survey*. Food Additives & Contaminants, 1999; 16(6): 253-260. DOI: 10.1080/026520399284019.
- Oveisi M-R, Jannat B, Sadeghi N, Hajimahmoodi M, Nikzad A. *Presence of aflatoxin M 1 in milk and infant milk products in Tehran, Iran*. Food Control. 2007; 18(10): 1216-8.
- Fallah AA. *Aflatoxin M 1 contamination in dairy products marketed in Iran during winter and summer*. Food control. 2010; 21(11): 1478-81. DOI: 10.1016/j.foodcont.2010.04.017.
- Rahimi E, Mohammadhosseini Anari M, Alimoradi M, Rezaei P, Arab M, Goudarzi M. *Aflatoxin M1 in pasteurized milk and white cheese in Ahvaz, Iran*. Global Veterinaria. 2012; 9(4): 384-7.

ACKNOWLEDGEMENTS

The authors would like to thank the Islamic Azad University, Mahabad Branch for supporting this study.

CONFLICT OF INTEREST

The authors have no conflict of interest to declare.

- Vagef R, Mahmoudi R. *Occurrence of Aflatoxin M1 in raw and pasteurized milk produced in west region of Iran (during summer and winter)*. International Food Research Journal. 2013;20(3): 1421-1425.
- Mahmoudi R, Golchin A, Hosseinzadeh N, Ghajarbeygi P. *Aflatoxin M1 and B1 contaminations in products of animal origin in Iran*. J Qazvin Univ Med Sci. 2014;18(4):49-59.
- Zinedine A, Juan C, Soriano J, Molto J, Idrissi L, Manes J. *Limited survey for the occurrence of aflatoxins in cereals and poultry feeds from Rabat, Morocco*. International Journal of Food Microbiology. 2007;115(1):124-7.
- Abdullah N, Nawawi A, Othman I. *Survey of fungal counts and natural occurrence of aflatoxins in Malaysian starch-based foods*. Mycopathologia. 1998;143(1): 53-8.
- Giray B, Girgin G, Engin AB, Aydın S, Sahin G. *Aflatoxin levels in wheat samples consumed in some regions of Turkey*. Food control. 2007;18(1): 23-9.
- Aydın A, Gunsen U, Demirel S. *Total aflatoxin, aflatoxin B1 and ochratoxin A levels in Turkish wheat flour*. Journal of Food and Drug Analysis. 2008; 16(2): 48-53.
- Taheri N, Semnani S, Roshandel G, Namjoo M, Keshavarzian H, Chogan A, et al. *Aflatoxin contamination in wheat flour samples from Golestan Province, Northeast of Iran*. Iranian journal of public health. 2012; 41(9): 42-47.
- Azizi G, Khoushnevis S, Hashemi S. *Aflatoxin M1 level in pasteurized and sterilized milk of Babol city*. Tehran University Medical Journal TUMS Publications. 2008; 65(13): 20-4.
- Halt M. *Aspergillus flavus and aflatoxin B1 in flour production*. European journal of epidemiology. 1994; 10(5): 555-8.
- Ayalew A, Fehrmann H, Lepschy J, Beck R, Abate D. *Natural occurrence of mycotoxins in staple cereals from Ethiopia*. Mycopathologia. 2006; 162(1): 57-63.
- Behfar A, Nazari Z, HEYDARI R. *Identification and Determination of Ochratoxin a Concentration in Wheat Flour of Flour Factories in the Ahvaz city Using HPLC*. Jundishapur Scientific Medical Journal. 2013; 12(2): 217-227.
- Yazdanpanah H, Miraglia M, Calfapietra F, Brera C. *Natural occurrence of aflatoxins and ochratoxin a in corn and barley from mazandaran and golestan in north provinces of IR Iran*. Mycotoxin research. 2001; 17(1): 21-30. DOI:10.1007/BF02946114.

25. Mahmoudi M, Aryaee P, Ghanbari M, Ansari H, Nourafcan H, editors. *The determination of aflatoxin and ochratoxin of flour and wheat in northern Iran*. International Conference on Environment, Agriculture and Food Sciences (ICEAFS'2012). 2012; 5-8.
26. Muscarella M, Palermo C, Rotunno T, Quaranta V, D'Antini P. *Survey of Ochratoxin A in cereals from Puglia and Basilicata*. Veterinary research communications. 2004; 28(Suppl 1): 229-32.
27. Baliukoniene V, Bakutis B, Stankevicius H. *Mycological and mycotoxicological evaluation of grain*. Annals of Agricultural and Environmental Medicine. 2003; 10(2): 223-7.
28. Basilio J. *Mycotoxins in food: PhD thesis*. National University of Litoral, Santa Fe, Argentina; 1995.
29. Obrian G, Georgianna D, Wilkinson J, Yu J, Abbas H, Bhatnagar D, et al. *The effect of elevated temperature on gene transcription and aflatoxin biosynthesis*. Mycologia. 2007; 99(2): 232-9.